Carbamazepine and CNV1427400 exhibit similar pharmacology in recombinant systems, rat and human native DRGs

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Introduction

Anticonvulsants decrease the hyperactivity of the pain network by targeting the ionic channels involved in the initiation and propagation of neuronal firing, such as voltage-gated sodium channels. However, current therapies are often associated with inconsistent efficacy and poor tolerability. In recent years there has been a huge effort from the pharmaceutical industry to develop new sodium channel blocking agents with improved pharmacology and safety profiles. It is therefore very important to confirm that the pharmacology and mechanism of action determined in recombinant human reagents or native rodent cells will translate to human native neurons. We have used whole cell patch-clamp electrophysiology to investigate the in vitro properties of Carbamazepine and CNV1427400, a novel sodium channel blocker, in human recombinant sodium channels, rat DRGs and human native DRGs from healthy donors.

Methods

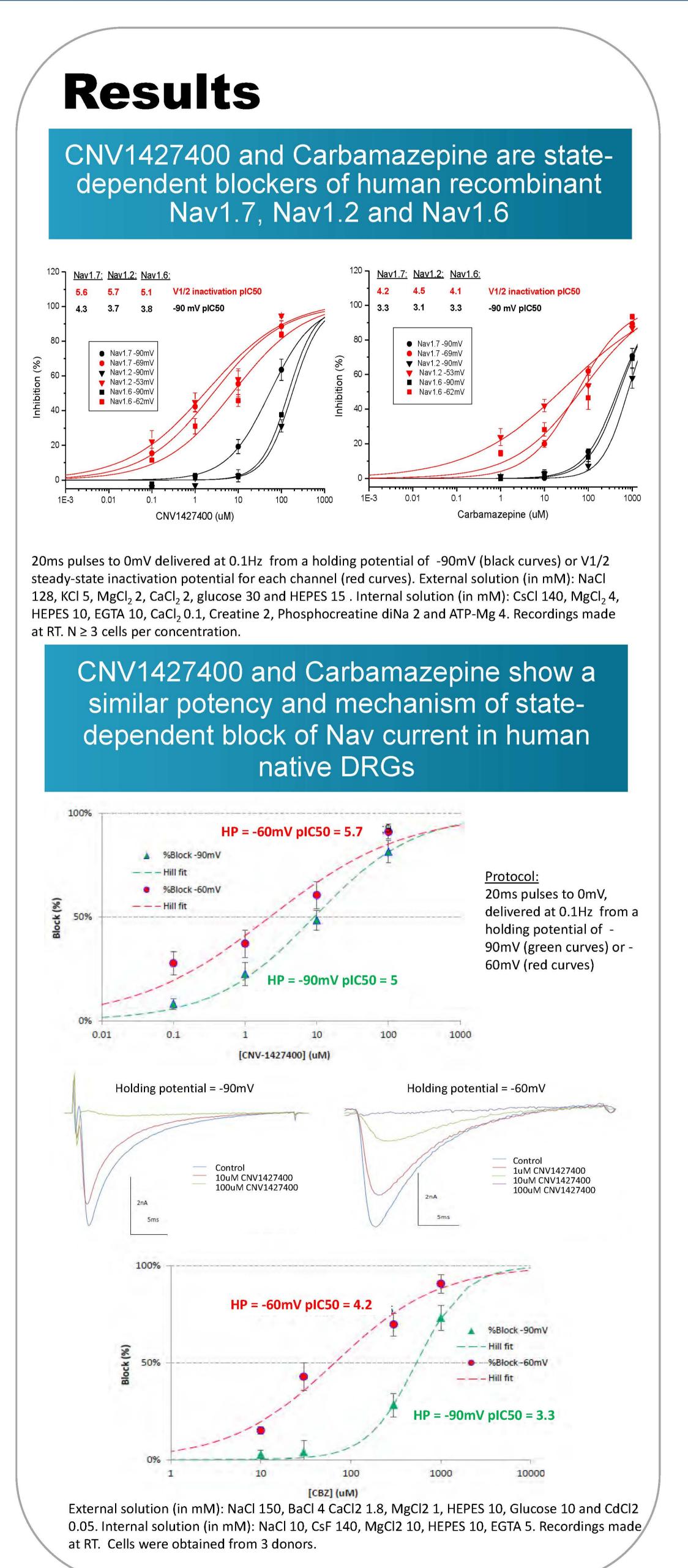
Rat DRGs:

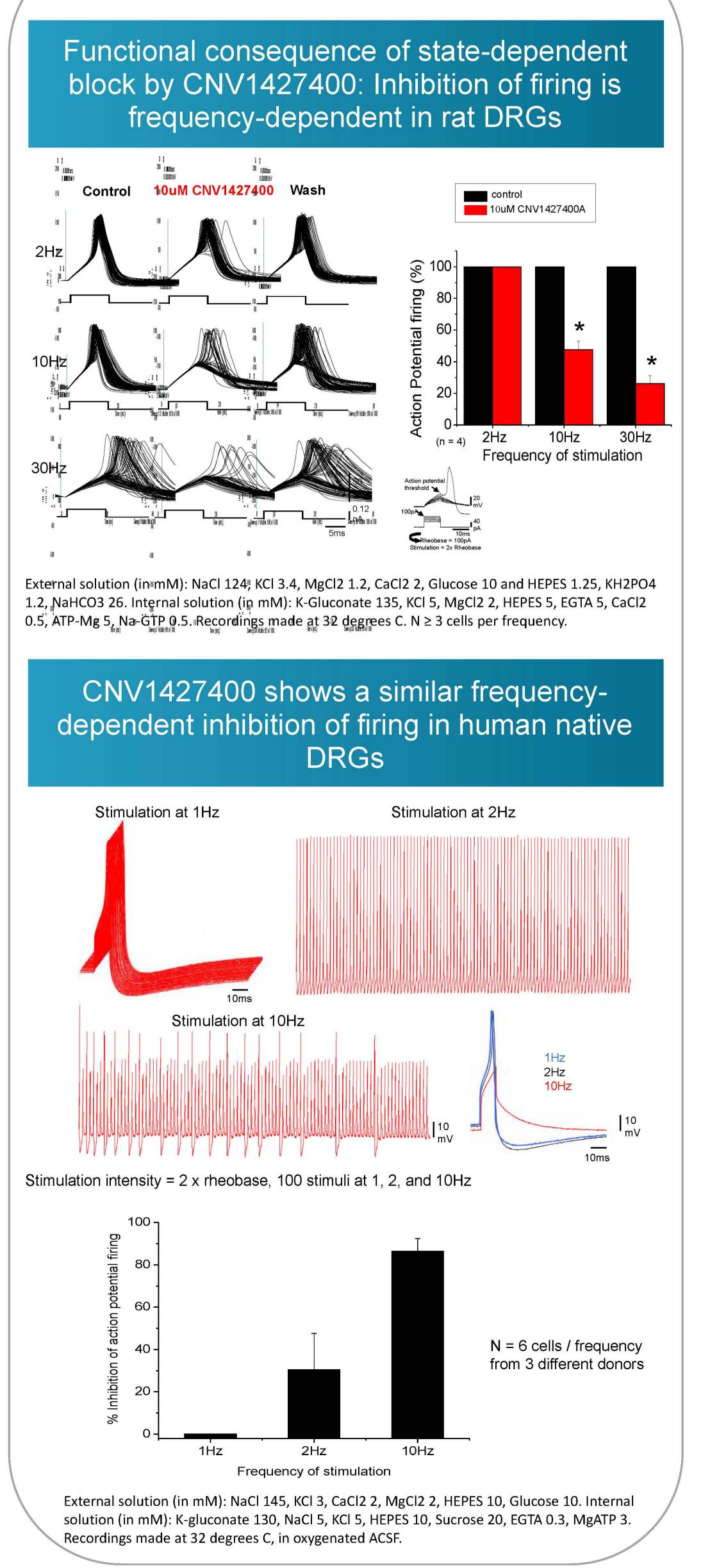
DRGs were obtained from neonatal rats (P7-10). Animals were euthanized in accordance with the 1986 Animals (Scientific Procedures) Act (Schedule 1). DRGs were dissected and dissociated by a prolonged collagenase treatment (0.1% in HBSS for 50 minutes at 37°C), followed by mechanical dissociation of the cells. They were then plated onto Laminin-coated coverslips (50,000 cells/ml).

Human DRGs:

All human tissues used for the study were obtained by legal consent from organ donors in the US. Donor DRGs were harvested using AnaBios proprietary surgical techniques. DRGs were then further dissected in cold neuroplegic solution to remove all connective tissue and fat. The ganglia were enzymatically digested and the isolated neurons put in culture in DMEM F-12 supplemented with Glutamine 2mM, FBS 10%, hNGF (10ng/mL), hGDNF (10ng/mL) and Penicillin / Streptomycin. Cells from 3 donors were used in this study.







CNV1427400 – broad efficacy across pain models and wide therapeutic index In vivo Model / Exposure Data behavioural test (unbound) Acute inflammatory pain MED 1mg/kg p.o. (*) 13ng/ml (ED₅₀ 0.6mg/kg p.o.) (intaplantar FCA) MED 1mg/kg p.o. (*), 10mg/kg p.o. (*), Chronic inflammatory pain (intra-articular FCA) equivalent to Celecoxib 30mg/kg p.o. Neuropathic pain (Chronic MED 0.5mg/kg p.o. (*) 9ng/ml Constriction Injury) equivalent to Gabapentin 30mg/kg p.o. Laboras (locomotor activity, NOAEL 40mg/kg p.o. 485ng/ml rearing and grooming) MED: minimum effective dose, NOAEL: no adverse event level, (*) statistical significance At doses showing efficacy in pain models, CNV1427400 inhibits WDR neuron firing in SNL rats baseline CNV-1427400A 0.3mg/kg CNV-1427400A 0.3mg/kg CNV-1427400A 3mg/kg CNV-1427400A 3mg/kg Recordings of WDR neurons from 7 CNV-1427400A 0.3mg/kg animals which received SNL (spinal CNV-1427400A 3mg/kg nerve ligation, Kim and Chung, 1992) surgery 14-18 days before use. All were confirmed as developing hindpaw hypersensitivity in behavioural testing at day 14 (Urch et al., 2003) • Compound administered s.c. at 0.3 or 3mg/kg, the mean plasma Cmax exposures were 37 and 417ng/ml respectively The effect of the compound was tested at 10, 30 and 50 minutes post Conclusions • In human native and recombinant Navs, Carbamazepine and CNV1427400 show similar potency and state-dependent mechanism of block, translating

into a similar frequency-dependent inhibition of action potential firing in rat

• Extending in vitro electrophysiology studies to human DRGs is a critical

technological breakthrough and adds great value from a translational point

of view when considering clinical dose prediction for the development of new

and human native DRGs.

analgesics.